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The Limits of Life and the Biosphere in Earth's Interior

By Verena B. Heuer, Mark A. Lever, Yuki Morono, and Andreas Teske

Fifty years of scientific ocean drilling have shown that microorganisms are widespread deep inside the ocean floor. Microbial populations exist in both organic-matter-rich and nutrient-poor sediments (Kallmeyer et al., 2012; D'Hondt et al., 2015), in sediments that are millions of years old and are buried to over a kilometer depth (Roussel et al., 2008; Ciobanu et al., 2014; Inagaki et al., 2015), and deep inside the basaltic oceanic crust (Orcutt et al., 2011; Lever et al., 2013). In these varied environments, metabolic activity is extraordinarily low (D'Hondt et al., 2009; Hoehler and Jørgensen 2013; Lever et al. 2015a), but microbial cells remain physiologically active (Morono et al., 2011) or survive in their dormant phases (Lomstein et al., 2012). The total amount of subsurface biomass is still being debated (Hinrichs and Inagaki, 2012; Kallmeyer et al., 2012; Parkes et al., 2014), and the factors posing ultimate limits to deep life

and the habitability of Earth remain to be resolved (Figure 1).

Within the ocean floor, geological, physical, chemical, and biological processes interact. Microorganisms can potentially exploit the energy released in these interactions, but they might also be substantially physiologically stressed in this environment. With depth below the seafloor, temperature increases and tectonic influences on heat and fluid flow result in vast temperature variations on local to global scales. Because temperature governs chemical reaction rates, its increase with depth becomes a major stress factor for microorganisms: rates of biomolecule-damaging reactions, such as DNA depurination, polymer hydrolysis, or amino acid racemization increase (Lindahl and Nyberg, 1972; Wolfenden et al., 1998; Steen et al., 2013), and so does the energy demand for biomolecule repair (Röling et al., 2003; Price and Sowers, 2004; Lever et al., 2015a).

Conversely, heating of sedimentary organic matter may also provide microbes with energy through the release of low-molecular weight compounds (Wellsbury et al., 1997; Horsfield et al., 2006). These temperature effects deserve attention as an estimated ~52% of Earth's marine sediment volume resides at temperatures >40°C, including ~25% at >80°C (LaRowe et al., 2017).

In deep, energy-limited subseafloor sediments, the upper temperature limit of life is expected to be lower than in surface hydrothermal habitats (Inagaki et al., 2015; Lever et al., 2015a; Møller et al., 2018), where abundant geothermally produced electron donors and seawater-derived electron acceptors provide energy for hydrothermal vent organisms to thrive. Indeed, pure cultures of certain hydrothermal vent archaea can be maintained at ~120°C under elevated pressure in the laboratory (Kashefi and Lovley, 2003; Takai et al., 2008), and several groups of Archaea and Bacteria from shallow seafloor environments thrive at temperatures of 80°–105°C both in the laboratory and in the field (e.g., Burggraf et al. 1990; Jørgensen et al., 1990, 1992; Lloyd et al., 2005; Edgcomb et al., 2007; Teske et al., 2009). By contrast, so far all attempts to isolate high-temperature-adapted microorganisms from deep subsurface sediments have failed. While the absence of microbial activity and viable microbial populations in deep oil reservoirs at temperatures above ~80°C (Wilhelms et al., 2001; Head et al., 2003) is consistent with known thermal limits

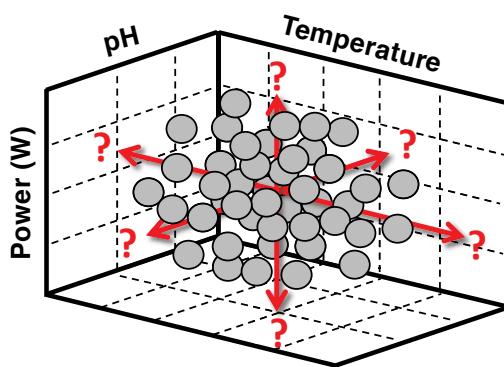


FIGURE 1. Concept sketch illustrating the niches of individual microbial strains (gray circles) with respect to power requirement (in Watts), pH range, and temperature range. The question marks indicate the currently unknown limits of microbial life in subsurface environments. For illustrational purposes, equal niche sizes among different microorganisms are assumed. In actuality, individual strains of subsurface microorganisms may differ greatly in cell-specific power, pH, and temperature ranges that allow them to thrive or survive. Furthermore, power requirements are likely to vary more systematically with pH and temperature than shown (i.e., subsurface microorganisms living at neutral pH and low temperature may generally have lower power requirements than counterparts living at extreme pH or temperature).

of microbial hydrocarbon degradation (Rueter et al., 1994; Holler et al., 2011; Kellermann et al., 2012; Laso-Perez et al., 2016; McKay et al., 2016), microbiological and geochemical investigations indicate even lower temperature maxima (~60°C) in deeply buried sediments (to ~2.5 km below seafloor; Inagaki et al., 2015).

Chemical disequilibria between seawater and crustal rocks (i.e., predominantly basalts) offer opportunities to exploit energy (e.g., Bach and Edwards, 2003), but the distribution and limits of life in subseafloor oceanic crust are even less understood than they are in sediments. Microbes in crustal rocks are difficult to sample with scientific ocean drilling because they are easily contaminated with microbes from drilling fluids. Sealed CORK borehole observatories, in which disturbances caused by drilling disappear and equilibrated fluid compositions similar to those in oceanic crust get

re-established within a few years (Wheat et al., 2010), are an important alternative to acquiring microbial samples from cores. CORKs provide access to fluid samples and free-living cells, and they can be used for colonization experiments (Cowen et al., 2003; Orcutt et al., 2011; Jungbluth et al., 2013).

The still-limited data from rocks and CORKs show large location-specific microbiological diversity, with only minor phylogenetic overlaps between free-living and attached cells (Biddle et al., 2014), and between microbes in crustal rocks and those inhabiting overlying sediments or seawater (Meyer et al., 2016; Labonté et al., 2017; Nigro et al., 2012). Moreover, microbial communities near cold recharge zones (Jørgensen and Zhao, 2016; Tully et al., 2018) differ distinctly from warm locations tens of kilometers downstream from recharge areas (Orcutt et al., 2010; Lever et al., 2013;

Robador et al., 2015). Yet, the factors controlling and limiting life in these hard rock environments are far from understood, and thus subseafloor oceanic crust remains a major frontier of deep biosphere research.

Ongoing analyses of samples collected by the International Ocean Discovery Program (IODP) promise to provide novel insights into some of these questions, because the challenges associated with accurate temperature measurement (Yanagawa et al., 2017), rigorous contamination control (e.g., Fisk et al., 2003; Lever et al., 2006, 2013; Santelli et al., 2010; Jørgensen and Zhao, 2016), and sensitive and reliable quantification of microbial populations and activity (Kallmeyer et al., 2008; Morono et al., 2013, 2014; Lever et al., 2015b; Glombitza et al., 2016; Ijiri et al., 2018) have been recognized and successfully addressed (Figure 2). Recent drilling into

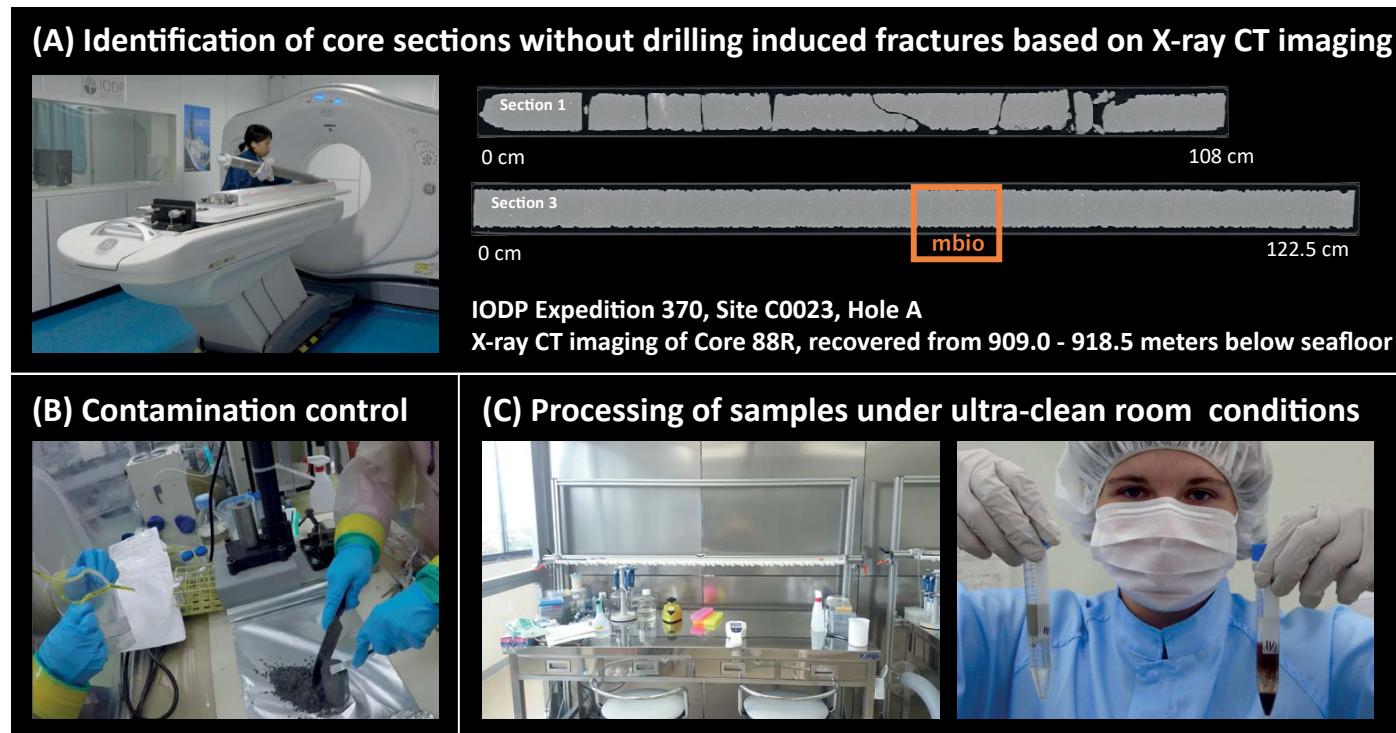


FIGURE 2. State-of-the-art contamination control for microbiological sampling from sediment cores during IODP Expedition 370. (A) On board the Japanese riser drilling vessel *Chikyu*, sediment cores were examined by X-ray computed tomography to identify lithological features and fractures induced in the course of drilling. Using the images, samples for microbiological investigations were selected from undisturbed core sections. The example shows the location of the “mbio” sample taken for cell counting. Photo credit: Dan Brinkhuis, SCIENCEMEDIA.NL, X-ray images from <http://sio7.jamstec.go.jp/xray-ct/370/C0023A/> (B) In order to avoid the intrusion of drilling fluid into the inner part of the “mbio” sample, the outer part was carefully removed with sterile tools inside an anaerobic chamber. (C) To avoid introduction of airborne contaminants into the sample, all further processing was conducted in an absolutely dust-free ultra-clean room laboratory at the Kochi Core Center/JAMSTEC. More detailed information is given in the Expedition Report (Heuer et al., 2016).

energy-poor subsurface sediment of the Nankai Trough subduction zone off Japan (IODP Expedition 370) and an upcoming expedition to energy-rich subsurface sediments of the Guaymas Basin in the Gulf of California (IODP Expedition 385, scheduled for 2019) will provide further constraints on the limits of life in subseafloor sedimentary environments as a function of energy supply and temperature-driven energy demand. Recent expeditions to the ultramafic Atlantis Massif on the Mid-Atlantic Ridge (IODP Expedition 357) and serpentinite mud volcanoes of the Mariana subduction zone (IODP Expedition 366) will offer new knowledge on the potential for deep water-rock reactions to fuel microbial life. Both of these locations also offer the opportunity to investigate the importance of variables other than temperature, for example, extreme pH, on the distribution of subseafloor microbial life. ☐

REFERENCES

- Bach, W., and K.J. Edwards. 2003. Iron and sulfide oxidation within the basaltic ocean crust: Implications for chemolithoautotrophic microbial biomass production. *Geochimica et Cosmochimica Acta* 67:3,871–3,887, [https://doi.org/10.1016/S0016-7037\(03\)00304-1](https://doi.org/10.1016/S0016-7037(03)00304-1).
- Biddle, J.F., S.P. Jungbluth, M.A. Lever, and M.S. Rappe. 2014. Life in the oceanic crust. Pp. 29–62 in *Microbial Life of the Deep Biosphere*. J. Kallmeyer and D. Wagner, eds, Walter De Gruyter GmbH, Berlin.
- Burggraf, S., H. Fricke, A. Neuner, J. Kristjansson, P. Rouvier, L. Mandelco, C.R. Woese, and K.O. Stetter. 1990. *Methanococcus igneus* sp. nov., a novel hyperthermophilic methanogen from a shallow submarine hydrothermal system. *Systematic and Applied Microbiology* 13:263–269, [https://doi.org/10.1016/S0723-2020\(81\)80197-9](https://doi.org/10.1016/S0723-2020(81)80197-9).
- Ciobanu, M.C., G. Burgaud, A. Dufresne, A. Breuker, V. Redou, S. Ben Maamar, F. Gaboyer, O. Vandenebeele-Trambouze, J.S. Lipp, A. Schippers, and others. 2014. Microorganisms persist at record depths in the subseafloor of the Canterbury Basin. *The ISME Journal* 8:1,370–1,380, <https://doi.org/10.1038/ismej.2013.250>.
- Cowen, J.P., S.J. Giovannoni, F. Kenig, H.P. Johnson, D. Butterfield, M.S. Rappe, M. Hutnak, and P. Lam. 2003. Fluids from aging ocean crust that support microbial life. *Science* 299:120–123, <https://doi.org/10.1126/science.1075653>.
- D'Hondt, S., F. Inagaki, C.A. Zarikian, L.J. Abrams, N. Dubois, T. Engelhardt, H. Evans, T. Ferdelman, B. Gribsholt, R.N. Harris, and others. 2015. Presence of oxygen and aerobic communities from sea floor to basement in deep-sea sediments. *Nature Geoscience* 8:299–304, <https://doi.org/10.1038/ngeo2387>.
- D'Hondt, S., A.J. Spivack, R. Pockalny, T.G. Ferdelman, J.P. Fischer, J. Kallmeyer, L.J. Abrams, D.C. Smith, D. Graham, F. Hasiuk, and others. 2009. Subseafloor sedimentary life in the South Pacific Gyre. *Proceedings of the National Academy of Sciences of the United States of America* 106:11,651–11,656, <https://doi.org/10.1073/pnas.0811793106>.
- Edgcomb, V.P., S.J. Molyneaux, S. Boer, C.O. Wirsén, M. Saito, M.S. Atkins, K. Lloyd, and A. Teske. 2007. Survival and growth of two heterotrophic hydrothermal vent archaea, *Pyrococcus* strain GB-D and *Thermococcus fumicola*, under low pH and high sulfide concentrations in combination with high temperature and pressure regimes. *Extremophiles* 11:329–342, <https://doi.org/10.1007/s00792-006-0043-0>.
- Fisk, M.R., M.C. Storrie-Lombardi, S. Douglas, R. Popa, G. McDonald, and C. Di Meo-Savoie. 2003. Evidence of biological activity in Hawaiian subsurface basalts. *Geochemistry, Geophysics, Geosystems* 4(12), <https://doi.org/10.1029/2002GC000387>.
- Glombitza, C., R.R. Adhikari, N. Riedinger, W.P. Gilhooley, K.U. Hinrichs, and F. Inagaki. 2016. Microbial sulfate reduction potential in coal-bearing sediments down to ~2.5 km below the seafloor off Shimokita Peninsula, Japan. *Frontiers in Microbiology*, <https://doi.org/10.3389/fmicb.2016.01576>.
- Head, I.M., D.M. Jones, and S.R. Larter. 2003. Biological activity in the deep subsurface and the origin of heavy oil. *Nature* 426:344–352, <https://doi.org/10.1038/nature02134>.
- Heuer, V.B., F. Inagaki, Y. Morono, Y. Kubo, L. Maeda, and the Expedition 370 Scientists. 2016. *Temperature Limit of the Deep Biosphere off Muroto*. Proceedings of the International Ocean Discovery Program, 370, International Ocean Discovery Program, College Station, TX, <https://doi.org/10.14379/iodp.proc.370.101.2017>.
- Hinrichs, K.-U., and F. Inagaki. 2012. Downsizing the deep biosphere. *Science* 338:204–205, <https://doi.org/10.1126/science.1229296>.
- Hoehler, T.M., and B.B. Jørgensen. 2013. Microbial life under extreme energy limitation. *Nature Reviews Microbiology* 11:83–94, <https://doi.org/10.1038/nrmicro2939>.
- Holler, T., G. Wegener, H. Niemann, C. Deusner, T.G. Ferdelman, A. Boetius, B. Brunner, and F. Widdel. 2011. Carbon and sulfur back flux during anaerobic microbial oxidation of methane and coupled sulfate reduction. *Proceedings of the National Academy of Sciences of the United States of America* 108:E1484–E1490, <https://doi.org/10.1073/pnas.1106032108>.
- Horsfield, B., H.J. Schenk, K. Zink, R. Ondrak, V. Dieckmann, J. Kallmeyer, K. Mangelsdorf, R. di Primio, H. Wilkes, R.J. Parkes, and others. 2006. Living microbial ecosystems within the active zone of catagenesis: Implications for feeding the deep biosphere. *Earth and Planetary Science Letters* 246:55–69, <https://doi.org/10.1016/j.epsl.2006.03.040>.
- Ijiri, A., F. Inagaki, Y. Kubo, R.R. Adhikari, S. Hattori, T. Hoshino, H. Imachi, S. Kawagucci, Y. Morono, Y. Ohtomo, and others. 2018. Deep-biosphere methane production stimulated by geofluids in the Nankai accretionary complex. *Science Advances* 4(6):eaao4631, <https://doi.org/10.1126/sciadv.aao4631>.
- Inagaki, F., K.-U. Hinrichs, Y. Kubo, M.W. Bowles, V.B. Heuer, W.-L. Hong, T. Hoshino, A. Ijiri, H. Imachi, M. Ito, and others. 2015. Exploring deep microbial life in coal-bearing sediment down to ~2.5 km below the ocean floor. *Science* 349:420–424, <https://doi.org/10.1126/science.aaa6882>.
- Jørgensen, B.B., M.F. Isaksen, and H.W. Jannasch. 1992. Bacterial sulfate reduction above 100°C in deep-sea hydrothermal vent sediments. *Science* 258:1,756–1,757, <https://doi.org/10.1126/science.258.5089.1756>.
- Jørgensen, B.B., L.X. Zawacki, and H.W. Jannasch. 1990. Thermophilic bacterial sulfate reduction in deep-sea sediments at the Guaymas Basin hydrothermal vent site (Gulf of California). *Deep Sea Research Part A* 37:695–710, [https://doi.org/10.1016/0198-0149\(90\)90099-H](https://doi.org/10.1016/0198-0149(90)90099-H).
- Jørgensen, S.L., and R. Zhao. 2016. Microbial inventory of deeply buried oceanic crust from a young ridge flank. *Frontiers in Microbiology*, <https://doi.org/10.3389/fmicb.2016.00820>.
- Jungbluth, S.P., J. Grote, H.T. Lin, J.P. Cowen, and M.S. Rappe. 2013. Microbial diversity within basement fluids of the sediment-buried Juan de Fuca Ridge flank. *The ISME Journal* 7:161–172, <https://doi.org/10.1038/ismej.2012.73>.
- Kallmeyer, J., R. Pockalny, R.R. Adhikari, D.C. Smith, and S. D'Hondt. 2012. Global distribution of microbial abundance and biomass in subseafloor sediment. *Proceedings of the National Academy of Sciences of the United States of America* 109:16,213–16,216, <https://doi.org/10.1073/pnas.1203849109>.
- Kallmeyer, J., D.C. Smith, A.J. Spivack, and S. D'Hondt. 2008. New cell extraction procedure applied to deep subsurface sediments. *Limnology and Oceanography—Methods* 6:236–245, <https://doi.org/10.4319/lom.2008.6.236>.
- Kashefi, K., and D.R. Lovley. 2003. Extending the upper temperature limit for life. *Science* 301:934, <https://doi.org/10.1126/science.1086823>.
- Kellermann, M.Y., G. Wegener, M. Elvert, M.Y. Yoshinaga, Y.-S. Lin, T. Holler, X.P. Mollar, K. Knittel, and K.-U. Hinrichs. 2012. Autotrophy as a predominant mode of carbon fixation in anaerobic methane-oxidizing microbial communities. *Proceedings of the National Academy of Sciences of the United States of America* 109:19,321–19,326, <https://doi.org/10.1073/pnas.1208795109>.
- Labonté, J.M., M.A. Lever, K.J. Edwards, and B.N. Orcutt. 2017. Influence of igneous basement on deep sediment microbial diversity on the eastern Juan de Fuca Ridge flank. *Frontiers in Microbiology*, <https://doi.org/10.3389/fmicb.2017.01434>.
- LaRowe, D.E., E. Burwicz, S. Arndt, A.W. Dale, and J.P. Amend. 2017. Temperature and volume of global marine sediments. *Geology* 45:275–278, <https://doi.org/10.1130/G38601.1>.
- Laso-Perez, R., G. Wegener, K. Knittel, F. Widdel, K.J. Harding, V. Krukenberg, D.V. Meier, M. Richter, H.E. Tegetmeyer, D. Riedel, and others. 2016. Thermophilic archaea activate butane via alkyl-coenzyme M formation. *Nature* 539:396–401, <https://doi.org/10.1038/nature20152>.
- Lever, M.A., M. Alperin, B. Engelen, F. Inagaki, S. Nakagawa, B.O. Steinsbu, and A. Teske. 2006. Trends in basalt and sediment core contamination during IODP Expedition 301. *Geomicrobiology Journal* 23:517–530, <https://doi.org/10.1080/01490450600897245>.
- Lever, M.A., K.L. Rogers, K.G. Lloyd, J. Overmann, B. Schink, R.K. Thauer, T.M. Hoehler, and B.B. Jørgensen. 2015a. Life under extreme energy limitation: A synthesis of laboratory- and field-based investigations. *FEMS Microbiology Review* 39:688–728, <https://doi.org/10.1093/femsre/fuv020>.
- Lever, M.A., O. Rouxel, J.C. Alt, N. Shimizu, S. Ono, R.M. Coggon, W.C. Shanks III, L. Lapham, M. Elvert, X. Prieto-Mollar, and others. 2013. Evidence for microbial carbon and sulfur cycling in deeply buried ridge flank basalt. *Science* 339:1,305–1,308, <https://doi.org/10.1126/science.1229240>.

- Lever, M.A., A. Torti, P. Eickenbusch, A.B. Michaud, T. Santi-Temkiv, and B.B. Jorgensen. 2015b. A modular method for the extraction of DNA and RNA, and the separation of DNA pools from diverse environmental sample types. *Frontiers in Microbiology*, <https://doi.org/10.3389/fmicb.2015.00476>.
- Lindahl, T., and B. Nyberg. 1972. Rate of depurination of native deoxyribonucleic acid. *Biochemistry* 11:3,610–3,618, <https://doi.org/10.1021/bi00769a018>.
- Lloyd, K.G., V.P. Edgcomb, S.J. Molyneaux, S. Boer, C.O. Wirsén, M.S. Atkins, and A. Teske. 2005. Effects of dissolved sulfide, pH, and temperature on growth and survival of marine hyperthermophilic archaea. *Applied and Environmental Microbiology* 71:6,383–6,387, <https://doi.org/10.1128/AEM.71.10.6383-6387.2005>.
- Lomstein, B.A., A.T. Langerhuus, S. D'Hondt, B.B. Jorgensen, and A.J. Spivack. 2012. Endospore abundance, microbial growth and necromass turnover in deep sub-seafloor sediment. *Nature* 484:101–104, <https://doi.org/10.1038/nature10905>.
- McKay, L., V.W. Klokman, H.P. Mendlovitz, D.E. LaRowe, D.R. Hoer, D. Albert, J.P. Amend, and A. Teske. 2016. Thermal and geochemical influences on microbial biogeography in the hydrothermal sediments of Guaymas Basin, Gulf of California. *Environmental Microbiology Reports* 8:150–161, <https://doi.org/10.1111/1758-2229.12365>.
- Meyer, J.L., U. Jaekel, B.J. Tully, B.T. Glazer, C.G. Wheat, H.T. Lin, C.C. Hsieh, J.P. Cowen, S.M. Hulme, P.R. Girguis, and J.A. Huber. 2016. A distinct and active bacterial community in cold oxygenated fluids circulating beneath the western flank of the Mid-Atlantic Ridge. *Scientific Reports* 6:22541, <https://doi.org/10.1038/srep22541>.
- Möller, M.H., C.G. Glombitsa, M.A. Lever, L. Deng, Y. Morono, F. Inagaki, M. Doll, C.-C. Su, and B.A. Lomstein. 2018. D:L-amino acid modeling reveals fast microbial turnover of days to months in the subsurface hydrothermal sediment of Guaymas Basin. *Frontiers in Microbiology* 9:967, <https://doi.org/10.3389/fmicb.2018.00967>.
- Morono, Y., T. Terada, T. Hoshino, and F. Inagaki. 2014. Hot-alkaline DNA extraction method for deep subseafloor archaeal communities. *Applied and Environmental Microbiology* 80:1,985–1,994, <https://doi.org/10.1128/AEM.04150-13>.
- Morono, Y., T. Terada, J. Kallmeyer, and F. Inagaki. 2013. An improved cell separation technique for marine subsurface sediments: Applications for high-throughput analysis using flow cytometry and cell sorting. *Environmental Microbiology* 15:2,841–2,849, <https://doi.org/10.1111/1462-2920.12153>.
- Morono, Y., T. Terada, M. Nishizawa, M. Ito, F. Hillion, N. Takahata, Y. Sano, and F. Inagaki. 2011. Carbon and nitrogen assimilation in deep subseafloor microbial cells. *Proceedings of the National Academy of Sciences of the United States of America* 108:18,295–18,300, <https://doi.org/10.1073/pnas.1007763108>.
- Nigro, L.M., K. Harris, B.N. Orcutt, A. Hyde, S. Clayton-Luce, K. Becker, and A. Teske. 2012. Microbial communities at the borehole observatory on the Costa Rica Rift flank (Ocean Drilling Program Hole 896A). *Frontiers in Microbiology*, <https://doi.org/10.3389/fmicb.2012.00232>.
- Orcutt, B.N., W. Bach, K. Becker, A.T. Fisher, M. Hentscher, B.M. Toner, C.G. Wheat, and K.J. Edwards. 2011. Colonization of subsurface microbial observatories deployed in young ocean crust. *The ISME Journal* 5:692–703, <https://doi.org/10.1038/ismej.2010.157>.
- Orcutt, B., C.G. Wheat, and K.J. Edwards. 2010. Subseafloor ocean crust microbial observatories: Development of FLOCS (FLow-through Osmo
- Colonization System) and evaluation of borehole construction materials. *Geomicrobiology Journal* 27:143–157, <https://doi.org/10.1080/01490450903456772>.
- Parkes, R.J., B. Cragg, E. Roussel, G. Webster, A. Weightman, and H. Sars. 2014. A review of prokaryotic populations and processes in sub-seafloor sediments, including biosphere:geosphere interactions. *Marine Geology* 352:409–425, <https://doi.org/10.1016/j.margeo.2014.02.009>.
- Price, P.B., and T. Sowers. 2004. Temperature dependence of metabolic rates for microbial growth, maintenance, and survival. *Proceedings of the National Academy of Sciences of the United States of America* 101:4,631–4,636, <https://doi.org/10.1073/pnas.0400522101>.
- Robador, A., S.P. Jungbluth, D.E. LaRowe, R.M. Bowers, M.S. Rappe, J.P. Amend, and J.P. Cowen. 2015. Activity and phylogenetic diversity of sulfate-reducing microorganisms in low-temperature subsurface fluids within the upper oceanic crust. *Frontiers in Microbiology* 5:748, <https://doi.org/10.3389/fmicb.2014.00748>.
- Röling, W.F.M., I.M. Head, and S.R. Larter. 2003. The microbiology of hydrocarbon degradation in subsurface petroleum reservoirs: Perspectives and prospects. *Research in Microbiology* 154:321–328, [https://doi.org/10.1016/S0923-2508\(03\)00086-X](https://doi.org/10.1016/S0923-2508(03)00086-X).
- Roussel, E.G., M.-A.C. Bonavita, J. Querellou, B.A. Cragg, G. Webster, D. Prieur, and R.J. Parkes. 2008. Extending the sub-sea-floor biosphere. *Science* 320:1,046, <https://doi.org/10.1126/science.1154545>.
- Rueter, P., R. Rabus, H. Wilkes, F. Aeckersberg, F.A. Rainey, H.W. Jannasch, and F. Widdel. 1994. Anaerobic oxidation of hydrocarbons in crude oil by new types of sulphate-reducing bacteria. *Nature* 372:455–458, <https://doi.org/10.1038/372455a0>.
- Santelli, C.M., N. Banerjee, W. Bach, and K.J. Edwards. 2010. Tapping the subsurface ocean crust biosphere: Low biomass and drilling-related contamination calls for improved quality controls. *Geomicrobiology Journal* 27:158–169, <https://doi.org/10.1080/01490450903456780>.
- Steen, A.D., B.B. Jorgensen, and B.A. Lomstein. 2013. Abiotic racemization kinetics of amino acids in marine sediments. *PLoS One* 10(4):e0123837, <https://doi.org/10.1371/journal.pone.0071648>.
- Takai, K., K. Nakamura, T. Toki, U. Tsunogai, M. Miyazaki, J. Miyazaki, H. Hirayama, S. Nakagawa, T. Nunoura, and K. Horikoshi. 2008. Cell proliferation at 122°C and isotopically heavy CH₄ production by a hyperthermophilic methanogen under high-pressure cultivation. *Proceedings of the National Academy of Sciences of the United States of America* 105:10,949–10,954, <https://doi.org/10.1073/pnas.0712334105>.
- Teske, A., V. Edgcomb, A.R. Rivers, J.R. Thompson, A.D. Gomez, S.J. Molyneaux, and C.O. Wirsén. 2009. A molecular and physiological survey of a diverse collection of hydrothermal vent Thermococcus and Pyrococcus isolates. *Extremophiles* 13:905–915, <https://doi.org/10.1007/s00792-009-0278-7>.
- Tully, B.J., C.G. Wheat, B.T. Glazer, and J.A. Huber. 2018. A dynamic microbial community with high functional redundancy inhabits the cold, oxic subseafloor aquifer. *The ISME Journal* 12:1–16, <https://doi.org/10.1038/ismej.2017.187>.
- Wellsbury, P., K. Goodman, T. Barth, B.A. Cragg, S.P. Barnes, and R.J. Parkes. 1997. Deep marine biosphere fuelled by increasing organic matter availability during burial and heating. *Nature* 388:573–576, <https://doi.org/10.1038/41544>.
- Wheat, C.G., H.W. Jannasch, A.T. Fisher, K. Becker, J. Sharkey, and S. Hulme. 2010. Subseafloor seawater-basalt-microbe reactions: Continuous sampling of borehole fluids in a ridge flank environment. *Geochemistry, Geophysics, Geosystems* 11(7), <https://doi.org/10.1029/2010GC003057>.
- Wilhelms, A., S.R. Larter, I. Head, P. Farrimond, R. di-Primio, and C. Zwach. 2001. Biodegradation of oil in uplifted basins prevented by deep-burial sterilization. *Nature* 411:1,034–1,037, <https://doi.org/10.1038/35082535>.
- Wolfenden, R., X.D. Lu, and G. Young. 1998. Spontaneous hydrolysis of glycosides. *Journal of the American Chemical Society* 120:6,814–6,815, <https://doi.org/10.1021/ja9813055>.
- Yanagawa, K., A. Ijiri, A. Breuker, S. Sakai, Y. Miyoshi, S. Kawagucci, T. Noguchi, M. Hirai, A. Schippers, J. Ishibashi, and others. 2017. Defining boundaries for the distribution of microbial communities beneath the sediment-buried, hydrothermally active seafloor. *The ISME Journal* 11:529–542, <https://doi.org/10.1038/ismej.2016.119>.

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